Pharmacological, bioactive screening of medicinal plant Nigella sativa and the derived compound thymoquinone: An invitro study

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ABSTRACT
The richest biosource of drugs are from the medicinal plants which are the traditional medicine. The nutraceuticals, food supplements, Siddha, Ayurveda, pharmaceutical medicines, synthetic drugs are from the plant sources. Inspite of large number of studies with herbal plants which have given good correlation in the phytochemical, anti-diabetic and anti-inflammatory content, Nigella sativa - a spice plant of Ranunculaceae family showed significant properties than their counterparts. The seeds of Nigella sativa and the essential oil were found to exhibit various pharmacological activities like antianalgesic, antiulcer, anti-inflammatory, antibacterial, antimicrobial, anticancer and anti-diabetic. Since there are no toxic effects or serious side effects observed using animal model and in the clinical trials, the study was carried out using Nigella sativa and Thymoquinone to find out the qualitative and quantitative phytochemical analysis, invitro - anti diabetic activity, anti inflammatory activity of Nigella sativa ethanolic extract and Thymoquinone effect on diabetes and Alzheimer’s disease. The aim of the study is to prove that the Nigella sativa could be used as therapeutic agent and the compound Thymoquinone a potential cholienesterase inhibitor for the treatment of Alzheimer’s disease.

INTRODUCTION
The richest biosource of drugs are from the medicinal plants which are the traditional medicine. The nutraceuticals, food supplements, Siddha, Ayurveda, pharmaceutical medicines, synthetic drugs are from the plant sources. Based on the senses conducted by the WHO 80-90% of drugs used by the people are originated from the traditional medicine (Arunkumar and Muthuselvam, 2009).

The drugs formulated are derived from the plants stem, bark, flowers, leaves, roots, seeds, fruits. To design the drug the phytochemical constituents are very important (Mojab et al., 2010; Parekh and Chanda, 2007). Similarly, various herbal extracts and essential oils are the richest source of phytochemicals such as tannins, carbohydrates, alkaloids, terpenoids, phenolics, flavonoids, steroids (Abbas et al., 2012). Inspite of large number of studies with herbal plants which have given good correlation in the phytochemical, anti-diabetic and anti-inflammatory content, Nigella sativa - a spice plant of Ranunculaceae family showed significant properties than their counterparts. The active compounds of Nigella sativa is used as a medicinal ailsments and Nigella sativa seeds are used as a medicinal plant for thousand years (Toncer and Kizil, 2004).

The seeds of Nigella sativa and the essential oil were found to exhibit various pharmacological activities like antianalgesic, antiulcer, anti-inflammatory, antibacterial, antimicrobial, anticancer and anti-
diabetic (Hanafy and Hatem, 1991). Nigella sativa has many active compounds including Thymoquinone, Longifolene, Flavone, Tetracosanoic acid, Acetin, oleic acid and linoleic acid. The seed oil is used as local anaesthetic (Warrier et al., 2004). Seeds and oil are the essential remedy for the treatment of various health issues including asthma, hypertension, diabetes, gastrointestinal problems, cough, jaundice, skin diseases, etc. Since there are no toxic effects or serious side effects observed using animal model and in the clinical trials, the study was carried out using Nigella sativa and Thymoquinone to find out the qualitative and quantitative phytochemical analysis, in vitro - anti diabetic activity, anti inflammatory activity of Nigella sativa ethanolic extract and Thymoquinone effect on diabetes and Alzheimer’s disease.

MATERIALS AND METHODS

Nigella sativa preparation

Nigella sativa seeds were purchased from local herbal store in Chennai. The seeds were authenticated and grounded to fine powder using electric grinder. The ground powder weighing 50 grams were soaked overnight in 150 ml of ethanol under shaking. The extracts were then filtered using whatmann no.1 filter paper and used the filtrate was used for further studies. The GCMS of Nigella sativa showed up various compounds out of which Thymoquinone (TQ) is used for further studies (A et al., 2020).

Figure 1: Qualitative analysis of Nigella sativa extract

Qualitative phytochemical analysis

Determination of total phenolic content

The determination of total phenolic content was studied using Folin-Ciocalteau (FC) method (Singleton et al., 1999). The reaction mixture was prepared using 0.2 ml gallic acid (standard), 1.0 ml of FC reagent, 0.8 ml of Na₂CO₃ solution were mixed well and allowed undisturbed for 2 hours at room temperature. The absorbance was measured using 750

Figure 2: DPPH radical scavenging activity of Thymoquinone

Figure 3: Hydrogen peroxide activity of Thymoquinone

Figure 4: AchE inhibitory Assay for Nigella sativa and Thymoquinone

Figure 5: Cytotoxicity of Thymoquinone

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Table 1: Phytochemical screening of Nigella sativa

<table>
<thead>
<tr>
<th>S.no</th>
<th>Phytochemical analysis</th>
<th>Nigella sativa extract</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td>Alkaloids</td>
<td>+</td>
</tr>
<tr>
<td>2.</td>
<td>Steroids</td>
<td>+</td>
</tr>
<tr>
<td>3.</td>
<td>Phenol</td>
<td>+</td>
</tr>
<tr>
<td>4.</td>
<td>Terpenoids</td>
<td>-</td>
</tr>
<tr>
<td>5.</td>
<td>Flavonoids</td>
<td>+</td>
</tr>
<tr>
<td>6.</td>
<td>Tannins</td>
<td>-</td>
</tr>
<tr>
<td>7.</td>
<td>Saponins</td>
<td>-</td>
</tr>
<tr>
<td>8.</td>
<td>Cardiac Glycosides</td>
<td>+</td>
</tr>
</tbody>
</table>

* + indicates the presence of the phytochemical constituents
- indicates the absence of the phytochemical constituents

Table 2: Metal chelating assay of Thymoquinone and Nigella sativa extract

<table>
<thead>
<tr>
<th>S.no</th>
<th>Parameters assessed</th>
<th>Thymoquinone</th>
<th>Nigella sativa</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td>Metal chelating assay</td>
<td>2.56±0.40</td>
<td>1.75±0.17</td>
</tr>
</tbody>
</table>

Figure 6: HRBC membrane stabilization assay of Thymoquinone

Figure 8: Alpha glucosidase inhibitory assay of Nigella sativa extract and Thymoquinone.

Figure 7: Alpha amylase inhibitory assay of Nigella sativa extract and Thymoquinone

Figure 9: Anti-inflammatory activity of Nigella sativa and Thymoquinone.

The total phenolic content (Ye) was expressed in terms as mg/ml.

Determination of total tannin content

Using Aluminium Chloride method (Ribereau-Gayon et al., 1998) the total tannin content (g/l) was determined. This method involves heating of tannin in acidic medium which leads to the formation of cyanidins. Two of the reaction tube contains, 1ml of sample, 0.5 ml water and 1.5 ml of 12 N HCl. The tube 1 was mixed and heated in water bath at 100°C for 30 minutes. The tube 2 is kept at room temperature. The tubes were cooled
and 0.25 ml of ethanol was added to the mixture and the absorbance was measured at 520 nm.

**Determination of total flavonoids**

The total flavonoids content was studied using (Zhuang et al., 2007). 1 ml of extract was added to 4 ml of water in 10 ml volumetric flask. After 5 mins of incubation, 5% of NaNO₂ (0.3 ml), 2% of AlCl₃ (1.5 ml) was added to the 10 ml volumetric flask. The mixture was then kept at constant shaking for 5 mins followed by the addition of 2 ml of NaOH (1M) with constant shaking. The absorbance was measured at 510 nm using blank. The content of total flavonoid was expressed in terms of mg of catechin equivalent / 1 g of content. The samples were analysed in duplicates and calculated based on the calibration curve of catechin.

**Phytochemical screening of Nigella sativa**

The crude extract of Nigella sativa was analysed to determine the bioactive components present in the extract as per the standard protocol described in (Hamburger and Hostettmann, 1991; Madhukar, 2013; Kamal, 2014).

**Test for alkaloids**

1.5 ml of 1% HCl was added to 2 ml of extract. The mixture was then heated in water bath followed by the addition of wanger’s reagent (6 drops). Orange precipitate formation indicates the presence of alkaloids.

**Test for steroids**

5 ml of chloroform and 2 ml of acetic anhydride was added to 2 ml of extract followed by the addition of concentrated sulphuric acid. Reddish brown colouration formed at the interface shows the presence of steroids.

**Test for phenol**

To 2 ml of extract, ferric chloride solution (5%) was added. The presence of phenol is identified by deep blue black colour.

**Test for terpenoids**

The extract is treated with chloroform along with few drops of concentrated sulphuric acid. The mixture is shaken well and allowed to stand undisturbed for few minutes. Yellow coloured layer formed at the lower region indicates the presence of terpenoids.

**Test for flavonoids**

To the extract ferric chloride solution was added in drops. The presence of flavonoid were detected using the intense green colour formed.

**Test for Tannins**

The extract was mixed with distilled water to make diluted test sample. To the sample 5% ferric chloride (2ml) was added. Blue green colour indicates the presence of tannins.

**Test for saponins**

The diluted extract was continuously shaken in a graduated cylinder for few mins. The foam layer which is formed on the top indicates the presence of saponins.

**Test for cardiac glycosides**

To the test sample (2ml), 3ml of glacial acetic acid followed by a drop of 5% ferric chloride was added. To the reaction mixture 0.5 ml of concentrated sulphuric acid was added along the sides of the test tube. The blue color formation in the acetic acid layer shows the presence of cardiac glycosides.

**Antioxidant activity**

DPPH (2,2 Diphenyl - 1 - picrylhydrazyl) radical scavenging activity of Thymoquinone

The compound of various concentration was added to DPPH (0.1mM). The DPPH alone serves as control. The thymoquinone was prepared using methanol. The samples were then incubated for half an hour in dark. The absorbance was measured 517 nm using UV - Vis spectrophotometry. Gallic acid was used as standard. The triplicates obtained was used to calculate the percentage of incubation (Gyamfi et al., 1999).

**Hydrogen peroxide scavenging activity of Thymoquinone**

The modified method of Dehpour was used to analyse and determine the scavenging activity of thymoquinone. Hydrogen peroxide (40mM) was prepared using phosphate buffer (pH 7.4) and various concentrations of compound was added and incubated for 30 mins. The absorbance was measured using UV.Vis spectrophotometry at 560 nm. Phosphate buffer alone serves as blank. Ascorbic acid was used as standard. The percentage of scavenging was calculated using the formula given below (Ngonda, 2013).

\[
\% \text{ scavenged} \ (H_2O_2) = 1 \times \frac{\text{Abs(std)}}{\text{Abs(cntrl)}} \times 100
\]

**Metal chelating assay**

500μM of freshly prepared FeSO₄ (150μL) was added to a mixture containing 0.1M of Tris - HCl (168μL), 0.8% w/v sodium chloride (218μL). To the mixture the extract of various concentration was added. 0.25% of 1,10-Phenanthroline (3μL) was
added to the mixture (Puntel et al., 2005). The absorbance was measured at 510 nm. The chelating activity of Fe$^{2+}$ was calculated using the equation:

\[
\%\text{Inhibition} = \left( \frac{\text{Abs}_{\text{control}} - \text{Abs}_{\text{sample}}}{\text{Abs}_{\text{control}}} \right) \times 100
\]

**Acetyl - cholinesterase activity assay**

The evaluation of acetylcholinesterase (AChE) activity of the extracts (Ellman et al., 1961). To each well of the microplate 40 microliter of enzyme (AChE) (0.28U/ml), 140μL of 5,5- dithiobis-(2-nitrobenzoic) acid (3.3mM) which was prepared in 0.1 M phosphate buffered solution of pH 7.0, which also contains 6mM of NaHCO$_3$. The extract of various concentration and 80 μL phosphate buffer of pH 8.0 was added to the mixture. The solution is incubated for about 20 mins at 25°C. Acetylthiocholine iodide (0.05mM) 40μL, was added to each well after the addition of substrate and the absorbance was measured at 412 nm. The enzyme inhibitory activity was expressed in terms of inhibition.

**Cytotoxicity assay**

The isolated lymphocyte cells were cultured in humidified 5% (v/v) CO$_2$ / air at 37°C in dulbecco’s modified eagle medium supplemented with 10% Fetal bovine serum and 100 U/ml penicillin. 5 x 10$^4$ cells/ml of cells were placed in 96 well plates. Insulin fibril was prepared by agitating the Insulin (Human recombinant insulin (10 mg/ml)) at 65°C for 5 hours (Jayamani and Shanmugam, 2014). The insulin fibril pre-formed with and without Thymoquinone was diluted with freshly prepared medium and added to the wells of final concentration 2μm oL/L. The same volume of medium is added to the control well. The plates were incubated at 37°C for 48 hrs. Cell viability was determined by using MTT toxicity assay. The MTT (5 mg/ml) was added to each well and incubated at 37°C for 3 hrs. The medium was removed and DMSO was added to each well. The plates were mixed well and the absorbance was measured at 490 nm using microplate reader (Khan et al., 2017).

**HRBC membrane stabilization assay of Thymoquinone**

Blood was drawn in a tube which contains EDTA which prevents coagulation. Various concentrations of the compound was added to 100μl of blood. Triton x 100, along with blood (positive control), and blood alone serves as negative control. The sample was incubated for 30 mins. The sample is then centrifuged at 3000 rpm for 20 mins and the supernatant was then removed for analysis. The absorbance was measured at 517 nm using UV-Vis spectrophotometer.

**Anti - diabetic activity of Nigella sativa extract and Thymoquinone**

\( \alpha \)-amylase inhibitory assay

According to (Zhang et al., 2010), the alpha amylose inhibitory studies were carried out. The Nigella sativa extract and the Thymoquinone compound were used for the studies. The various concentrations of the extracts and the compound was mixed with di-methyl sulfoxide solution. The alpha amylose was dissolved in phosphate buffer of pH 6.8. After incubation, starch solution (1%) was added to all the tubes. The reaction mixture was incubated again for 15 minutes. After the incubation time the reaction was stopped using 1 ml of di-nitro salicylic acid (DNS) reagent and the tubes were boiled in water bath for about 10 minutes. The contents were then cooled, to which 10 ml of distilled water was added. Absorbance was measured at 540 nm. Acarbose serves to be the positive control.

\[
\text{Percentage of inhibition} = \left( \frac{\text{Abs}_{\text{control}} - \text{Abs}_{\text{test sample}}}{\text{Abs}_{\text{control}}} \right) \times 100
\]

\( \alpha \)-glucosidase inhibitory assay

\( \alpha \)-glucosidase inhibitory assay was determined based on the protocol (Ting et al., 2005). 112μL of potassium phosphate buffer (pH 6.8), 20 μL of enzyme solution, 8μL of the extract and compound TQ were mixed and incubated at 37°C for 15 minutes. Following 20 μL of p-NPG was added and further incubated for another 15 minutes at 37°C. The reaction was terminated with 80μL of Na$_2$CO$_3$ solution. Absorbance was measured at 405 nm. 8μL of DMSO was added to control and blank incase of extract and compound. Acarbose serves as positive control.

\[
\text{Inhibitory activity} (%) = \left[ 1 - \left( \frac{\text{OD}_{\text{sample}} - \text{OD}_{\text{sample blank}}}{\text{OD}_{\text{control}} - \text{OD}_{\text{blank}}} \right) \right] \times 100
\]

**Anti inflammatory activity of Nigella sativa extract Thymoquinone**

Interleukin -1 beta (IL-1β) were used to analyse using lymphocyte cell culture supernatant using ELISA kit which is available commercially. Based on the bioassay technology protocol the lymphocyte sample was processed and the triplicates were obtained (Quillin et al., 2006).

**RESULTS AND DISCUSSION**

Qualitative phytochemical analysis of Nigella sativa extract
The Figure 1, shows the highest concentration of total phenolic content was recorded in the ethanolic extract of Nigella sativa. The amount of polyphenols in ethanolic extract was 35.6 mg/ml. Similarly, in flavonoids and tannins it is obtained as 1.5 mg/ml and 2.6 mg/ml. These results are based on the study made by (Cheikh-Rouhou et al., 2007) which states that Nigella sativa is the potential source of phenolic compounds.

**Phytochemical screening of Nigella sativa**

In the analysed results it is evident that the phytochemical screening of the Nigella sativa extract showed the presence of secondary metabolities like Alkaloids, Steroids, Flavonoids, Glycosides (Table 1). These compounds are with medicinal as well as physiological benefits. This isolation of bioactive compounds helps in the formulation of new drugs for various diseases and disorders.

**Anti - oxidant activity**

**DPPH radical scavenging activity of Thymoquinone**

DPPH radicals are reduced due to the donation of hydrogen molecules which is more stable and estimates free radicals. The DPPH activity of the Thymoquinone (IC 50 value 121.56±0.66μg/ml) is showed in the Figure 2. The DMSO dissolved compound thymoquinone showed low scavenging activity when compared with gallic acid (IC 50 value = 2.46±0.76μg/ml). Nigella sativa extract exhibited IC 50 value 11.26±0.93 μg/ml. Based on this result it is evident that thymoquinone has a low potential effect towards the antioxidant activity whereas Nigella sativa exhibited good potential towards antioxidant activity (A et al., 2020).

**Hydrogen peroxide scavenging activity of Thymoquinone**

The compound Thymoquinone was analyzed by measuring decrement of hydrogen peroxide at 230 nm. The compound is found to be more effective whose IC 50 value is 12.31 μg/ml and dose dependent, as similar to Nigella sativa (A et al., 2020) when compared to the standards ascorbic acid whose IC 50 value is 17.50μg/ml, Figure 3.

**Metal chelating assay of Thymoquinone and Nigella sativa extract**

As shown in the results the extract Nigella sativa chelates the metal Fe(II) at various concentrations whose mean±SD values are found to be 1.75±0.17 when compared to thymoquinone (2.56±0.40).The result revealed that the extract has high scavenging ability of OH when compared to TQ which is less efficient in scavenging the OH molecules. (Table 2)

**Acetyl - Cholinesterase activity of Thymoquinone and Nigella sativa extract**

Cholinesterase study reveals the role of TQ as a cholinesterase inhibitor. The inhibition of the enzyme cholinesterase by TQ showed higher inhibition when compared to Nigella sativa extract. The highest cholinesterase activity of TQ was 53.7 ±1.1 μg/ml, in which Nigella sativa extract exhibited 84.7 ±4.3μg/ml. This indicates that the TQ is effective in preventing the enzyme cholinesterase’s role in neurotransmitter AchE hydrolysis (Figure 4).

**Cytotoxicity assay of Thymoquinone**

The isolated lymphocytes were treated with Thymoquinone (TQ) compound to study the cytotoxicity of the cell. The assay showed statistical decrease in cell death at the concentration of 100μg (40%) when compared with the control. This showed a less effect on cell proliferation. (Figure 5).

**HRBC membrane stabilization assay of Thymoquinone**

The hemolysis prevention using Thymoquinone compound is determined using the stabilization studies. Triton x 100 destabilizes the RBC membrane and makes the hemoglobin to leak. Different concentrations of the compound were analysed and it was found that Thymoquinone prevents the leak of hemoglobin from the RBC by stabilizing the membrane. This even suggests that Thymoquinone is non toxic and it can be used for biological studies Figure 6.

**Anti - diabetic activity of Thymoquinone and Nigella sativa extract**

**α- amylase inhibitory assay**

The alpha amylase inhibitory assay of Nigella sativa showed the highest inhibition rate of about 50.84% at 100 μg/ml concentration whereas for TQ the highest inhibition rate 64.09% at 100 μg/ml concentration and the lowest inhibition rate of about 15.49 % 10 μg/ml concentration while TQ showed inhibition rate of 12.02% at 10 μg/ml concentration as shown in Figure 7. Acarbose serves as positive control. The result showed that the extract and the compound TQ is dose dependent and revealed the increase in inhibitory activity against amylase enzyme.TQ showed potential activity towards inhibition when compared to Nigella sativa extract.

**α- glucosidase inhibitory assay**

Alpha glucosidase inhibitory activity of Nigella sativa extract and TQ was used to analyse the inhibitory activity of the samples. The inhibitory activity was determined by using p-Nitrophenyl - α - D- glucopyranoside (p-NPG) which is a substrate
and they are compared with the control Acarbose. The extract of concentration 5, 15, 25, 50 μg/ml showed less than 50% of inhibition whereas 100 μg/ml showed more than 50% of inhibition when compared to TQ. TQ inhibited more than 50% at concentrations 50 and 100 μg/ml. Based on the results, TQ showed a effective inhibitory activity when compared with the extract Figure 8.

**Anti-inflammatory activity of Thymoquinone and Nigella sativa extract**

Increased level of IL-1β cytokine is the major cause in various metabolic disorders. The inhibition of these will prevent the pathogenic effects which leads to metabolic disorders (Bang et al., 2009). The down regulation of these is an important goal in pharmacology for the inflammatory related disorders. The extract and compound TQ for the inhibition of IL-1β were analysed and the effect was expressed in Figure 9. It is evident that TQ is more potent than extract in the inhibition of IL-1β. This is compared with the standard IL-1β, and also with hydrogen peroxide treated cells. Based on the results the TQ showed inhibition, it has a potent anti-inflammatory property.

**CONCLUSIONS**

The invitro studies revealed that the crude extract of Nigella sativa have many phytochemical constituents that are medicinally significant. Crude extract of Nigella sativa and the isolated compound Thymoquinone, have anti-inflammatory and anti-diabetic activities which was evident from IL-1 β assay, Alpha amylase and alpha glucosidase inhibitory assay. Though Nigella sativa is more effective in DPPH radical scavenging and metal chelating assay than Thymoquinone, the latter (TQ) showed more efficacy in hydrogen peroxide scavenging activity and AchE inhibitory assay. Such properties make Thymoquinone a potential and promising compound in the treatment of Alzheimer’s disease.

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