



# INTERNATIONAL JOURNAL OF RESEARCH IN PHARMACEUTICAL SCIENCES

Published by Pharmascope Publications

Journal Home Page: [www.pharmascope.org/ijrps](http://www.pharmascope.org/ijrps)

## Open source software tools for computer aided drug design

Gurtej Kanwar<sup>1</sup>, Anish Kumar\*<sup>1</sup>, Anshika Mahajan<sup>2</sup><sup>1</sup>School of Bioengineering and Biosciences, Lovely Professional University, Punjab, India<sup>2</sup>Department of Chemistry, Jammu University, Jammu, India

### Article History:

Received on: 21.12.2017  
 Revised on: 11.02.2018  
 Accepted on: 12.02.2018

### Keywords:

Molecular docking  
 Energy minimization  
 Structure refinement  
 Drug design  
 CADD

### ABSTRACT

Computer-aided drug design (CADD) has revolutionized the drug discovery arena and it has reduced the costs associated with finding novel compounds which are having pharmaceutical importance. In CADD, the scientists use the computer software to discover biological active compounds. Molecular docking and energy minimization tools are essential components of structure based drug design. It is a significant tool in structural molecular biology and computer-assisted drug design. It reduces the laboratory workload of the end user and allows researchers to restrict their docking studies to the smallest and the most representative set of macromolecules and small molecules possible. This greatly enhances the productivity of researchers. Energy minimization is an important criterion for selecting a potential 3D molecule. In modeled structures, the 3D structure is affected is due to steric clashes. These clashes happen in a protein structure due to the overlap of non bonding atoms and with the assistance of energy minimization, steric clashes can be eradicated. The open software's and databases provides a platform for scientists and scholars to carry out their research work in a better way. The docking tools are discussed in this review cover protein-ligand, protein-peptide as well as protein-nucleic acid docking. The tools described include AutoDock 4 and Vina, UCSF DOCK, FLIPdock, EADock, HADDOCK 2.2, SwissDock, PatchDock and ClusPro. In addition to the docking tools, energy minimization tools such as YASARA minimization server, KoBaMIN server and 3D refine server have also been discussed. This mini-review concentrates on open software tools which are free of cost and can be easily downloaded in the computers that are useful for CADD.



### \* Corresponding Author

Name: Anish Kumar  
 Phone: +91-  
 Email: anish.20215@lpu.co.in

ISSN: 0975-7538

Peer reviewed

DOI: <https://doi.org/10.26452/ijrps.v9i1.1179>

Production and Hosted by

Pharmascope.org

© 2018 Pharmascope Publications. All rights reserved.

### INTRODUCTION

Computer aided drug design has played a key role in drug discovery from the past thirty years. Molecular docking software's are an integral part of any structure based drug design process. They predict the formation of non-covalent bond between a ligand (usually a small molecule) and a macromolecule (usually a receptor protein) (Trott *et al.*, 2010). In structure based drug design, the ligands are usually the small molecule drug candidates whose non-covalent interactions with a target receptor protein are to be simulated computationally. Several docking tools are available at no cost to the end user, allowing unrestricted access to carry out virtual high throughput screening (VHTS) of several ligands at once in even resource constrained laboratories such as those in several

developing countries. Virtual screening has expedited and reduced the cost of drug discovery process (Kumar *et al.*, 2016). Various kinds of kinase have been discovered with the help of virtual screening. The most commonly cited are human immunodeficiency virus (HIV) drugs, such as amprevir (Agenerase) and nelfinavir (Viracept), which were developed using the crystal structure of HIV protease. This methodology gets to be main stream in the pharmaceutical examination for lead molecule classification. It is envisaged as substitute path for trial screening of drug molecules. It demonstrates an expanded achievement rate in the process of drug findings. It allows rapid and inexpensive filtering of active compounds from inactive ones. As a result, virtual screening has become an essential part of modern computer aided drug discovery (CADD) process. Databases such as AfrodDb, iSMART, Traditional Chinese Medicine (TCM), Super Natural II, PubChem, ZINC database etc are a useful source of ligands for carrying out virtual screening (McInnes, 2007). Although docking servers for protein-protein and protein-small molecule docking are widely available, servers for protein-nucleic acid docking have so far been relatively few. Most of the servers which do allow protein-nucleic acid docking were initially developed for protein-protein docking and later modified so as to accommodate protein-nucleic acid docking (Tuszynska *et al.*, 2015). This, however, is likely to change soon as a result of the growth of interest in ncRNA (non-coding RNA) due to their role in disease, development and the potential of exogenous ncRNA (such as siRNA) in therapeutics. In addition to protein-nucleic acid docking tools, siRNA design tools are also likely to benefit due to the increased interest in the role of RNAi (RNA interference) in the regulation of gene expression in higher organisms (Laganà *et al.*, 2015).

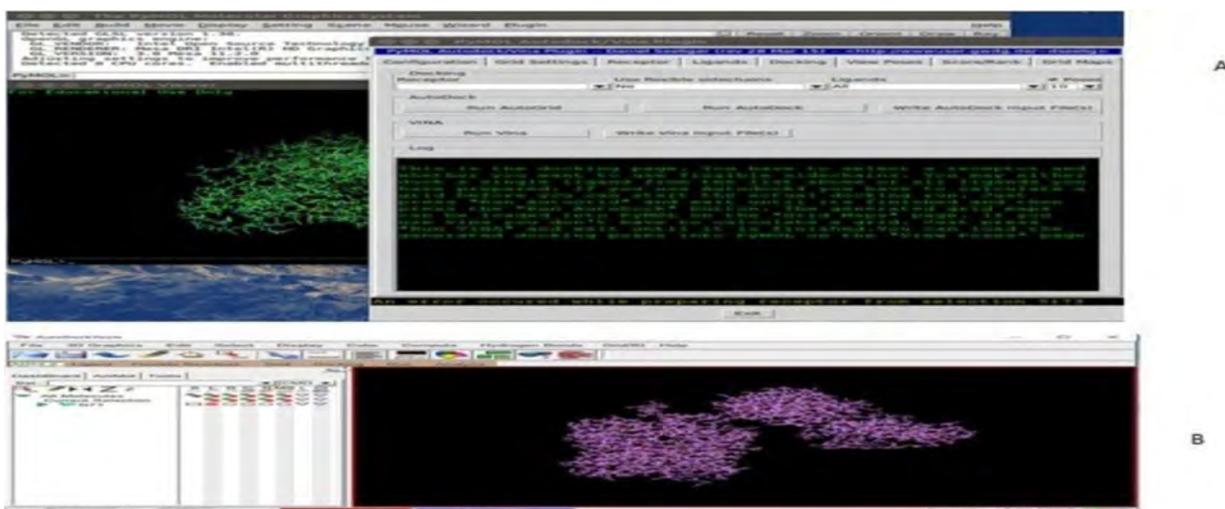
Energy minimization tools (also called structure refinement tools) are another important component of a structure based drug design workflow. Software programs are employed to create the 3D structures. After the 3D structure is built, energy minimization is carried out since it results in unfavorable bonded and non-bonded interactions. These clashes happen in a protein structure due to the overlap of non bonding atoms and with the assistance of energy minimization, steric clashes can be eradicated (Ramachandran *et al.*, 2011). Energy minimization is done to bring the potential energy of the system to the lowest point and to eliminate close contacts. This enables the user to find out the structure with the minimum potential energy (and hence maximum stability) from a given set of atomic coordinates and interatomic bonds. In this review article, we have summarized some of the important protein structure refinement tools are

that inexpensive and most of the popular structure refinement tools are web based.

### Molecular Docking Tools

Molecular docking software predicts the non-covalent interactions occurring between ligands and their corresponding binding sites in the receptors (Kumar *et al.*, 2015; Kumar *et al.*, 2017). Depending on whether the binding site must be defined before the docking run, the docking can be classified as either blind docking (binding site not defined) or local docking (binding site is defined). Most modern molecular docking software carry out local docking as blind docking is much more expensive computationally. The only case in which blind docking might be considered plausible is when the target protein is small in size. Another way of classifying docking is on the basis of whether the ligand or the target of interest is flexible. On the basis of flexibility, docking can be classified as being either rigid or flexible (Teague, 2003). Flexible docking can be further classified as flexible ligand-rigid protein, flexible ligand-flexible protein and rigid ligand ligand-flexible protein. Flexible proteins, although more computationally expensive to consider than rigid proteins, greatly improve the accuracy of the results (Carlson and McCammon, 2000; Kumar and Ramanathan, 2014). As a result of the computationally intensive nature of docking, docking servers have gained widespread popularity in the scientific community as it provides the end user access to powerful hardware via an easy to use web interface. ClusPro server was the first molecular docking server (Comeau *et al.*, 2005).

The structure of the protein on which the docking run is being performed is in most cases obtained by X-ray crystallography. NMR (Nuclear Magnetic Resonance) may also be used in some cases, especially when the protein cannot be crystallized. In most structure based drug design scenarios, the set of ligands under consideration are the possible drug candidates and the target a receptor or an enzyme. However, this is only one of the several possible intermolecular interactions amenable to study using molecular docking tools (Meng *et al.*, 2011). Due to growing interest in RNA based therapeutics in the pharmaceutical industry, there is an increasing requirement of molecular docking tools which can predict protein-nucleic acid and nucleic acid-nucleic acid interactions. The main aim of molecular docking tools is to determine the docking pose (a docking pose is the conformation of ligand and target molecules at the time of binding) which has the minimum free energy of binding. Any good docking tool must have a good accuracy in predicting ligand-target interactions and must maximize



**Figure 1: Snapshot showing (A) Autodock/Vina plugin for PyMOL and (B) AutoDockTools, the GUI for AutoDock/Vina**

WELCOME TO THE UTRECHT BIOMOLECULAR INTERACTION WEB PORTAL >>

This is the easy interface to the HADDOCK docking program. Please define the structure for each molecule you want to dock as well as the residues belonging to the interaction interface.  
 Docking is performed with default settings that work well for average complexes. If you do not have any special wishes for the system you want to have docked, this is the way to go.  
 Unfold the menus by clicking on the double arrows; Submit your job by providing your username and password and press submit.  
 For questions about the use of the HADDOCK portal please refer to: ask.bioexcel.eu

You may supply a name for your docking run (one word)  
 Name

**First molecule**

**Structure definition**  
 Where is the structure provided?   
 Which chain of the structure must be used?   
 PDB structure to submit  Browse... No file selected.  
 or: PDB code to download

**Restraint definition**  
 Data to drive the docking  
 Please supply residues as comma-separated lists of residue numbers  
 Active residues (directly involved in the interaction)   
 Passive residues (surrounding surface residues)   
 Define passive residues automatically around the active residues   
 What kind of molecule are you docking? Protein/peptide/ligand

**Second molecule**

**Structure definition**  
 Where is the structure provided?   
 Which chain of the structure must be used?   
 PDB structure to submit  Browse... No file selected.  
 or: PDB code to download

**Restraint definition**  
 Data to drive the docking  
 Please supply residues as comma-separated lists of residue numbers  
 Active residues (directly involved in the interaction)   
 Passive residues (surrounding surface residues)   
 Define passive residues automatically around the active residues   
 What kind of molecule are you docking? Protein/peptide/ligand

**Figure 2: Snapshot showing HADDOCK 2.2 web server interface**

its computational speed for a given set of parameters (Kumar *et al.*, 2016).

Docking tools are highly useful for rapid and efficient virtual screening of several candidate drug molecules. In virtual screening, several ligands are tested in various conformations in the binding site of their targets and the corresponding free energies of binding determined. The conformations with the lowest free energy of binding are chosen. This enables the researcher to filter out and identify suitable lead compounds to work on from a

large set of candidate drug molecules. The two most important features of a docking software is the scoring function it uses for the ligand-receptor complex and the algorithm used for finding conformations of ligand in binding site of the receptor. Scoring functions allow the docking program to rank the affinity between the ligand and the binding site in the receptor. A good scoring function should strike an adequate balance between efficiency in usage of computational resources and accuracy in ranking affinities (Forli *et al.*, 2016).

### AutoDock 4 and AutoDock Vina

AutoDock 4 and AutoDock Vina are popular molecular docking programs developed at The Scripps Research Institute. These two software are not only free but also open source, allowing the end user to make improvements and modifications in the underlying source code. The key difference between AutoDock 4 and Vina is in their scoring functions. Although the base installation of AutoDock 4 and Vina provides access to only their command line interface, a graphical user interface, AutoDockTools (ADT), can be downloaded separately as part of the MGLTools software. Alternatively, PyMol plugin autodock.py can also be used to view docking poses generated by AutoDock 4 and Vina (Chaitanya *et al.*, 2010). This gives the user access to the viewing capabilities of PyMol and docking capabilities of AutoDock 4 and Vina. After a docking run has been completed, the docking scores of various poses can be exported in diverse formats. AutoDock utilizes the Lamarckian Genetic algorithm (LGA) during the docking run (Seeliger *et al.*, 2010; Kumar *et al.*, 2014).

### UCSF DOCK

UCSF DOCK was one of the first molecular docking software. It assumed both the receptor and the ligand to be rigid initially (Clark and Ajay, 1995). This is the least computationally intensive way of carrying out a docking run. However, the results in this case are far from accurate. As a result, as computational power increased, flexible ligand docking was incorporated in later versions of DOCK. DOCK works mainly by superimposing the ligand onto a negative image of the binding site of the receptor. It screens large libraries of small molecules which could serve as potential ligands to determine those that fit the binding site the 'best' (Kolb *et al.*, 2009). The latest version of DOCK is DOCK 6. A highly useful feature of DOCK 6, especially due to increasing interest in RNA therapeutics, is the ability of DOCK 6 to be used for nucleic acid targets in addition to the protein targets (Lang *et al.*, 2009). The versatility of DOCK is demonstrated by the fact that Hermann *et al.* utilized DOCK for structure based prediction of function of enzymes. This involved the docking of high energy intermediates to the active site of enzymes. However, there are limitations to such applications of DOCK. Enzymes can undergo significant changes during the course of a reaction. Additionally, when utilizing DOCK for such applications, only a limited set of substrates can be considered (Hermann *et al.*, 2007).

### HADDOCK 2.2

HADDOCK 2.2 (High Ambiguity Driven Protein-Protein Docking) web server primarily is meant for

protein-protein and protein-peptide docking simulations. It was originally developed for NMR data. It has a large user base in India. As of 2016, HADDOCK server has had over 6000 users and has completed more than 108,000 runs. More than 120 protein structures, whose structure have been calculated using HADDOCK, have been submitted in PDB. HADDOCK server gives access to seven interfaces to the user. The interfaces differ in the number of parameters that can be changed. The most basic interface is the 'Easy' interface. The 'Guru' interface is the most advanced interface, allowing access to all the molecular docking parameters available on HADDOCK web server. 'Guru' and 'Expert' allow access to advanced level features such as the ability to choose which region of the molecule are to be considered flexible or semi-flexible (Van *et al.*, 2016). Since docking calculations requiring access to advanced level features are computationally much more expensive than those that can simply be carried out by 'Easy' interface options, access to advanced level interfaces is only granted upon request. Users can send the request to [haddock@ gmail.com](mailto:haddock@ gmail.com). 'Easy' and 'Prediction' interfaces can be used without requesting access. Upon completion of docking run, the user would receive an email containing a link to the results page. The HADDOCK score displayed on the results page takes into account the Vander Waals forces, electrostatic forces, desolvation energy, restraint violation energy and buried surface area at the region of interaction between the interacting molecules. Unlike most other docking tools, HADDOCK also has the capability to deal with more than 2 molecules simultaneously per docking run (Karaca *et al.*, 2010). The interface for access to multiple molecule docking features is the 'Multi-body interface' of the HADDOCK web server. Although HADDOCK is primarily used for protein-protein and protein-peptide docking studies, HADDOCK versions 2.0 and onward also support nucleic acid and small molecule docking (Vries *et al.*, 2010).

### PatchDock

PatchDock web server, which runs on PatchDock algorithm, is useful for protein-small molecule and protein-protein docking (Kumar *et al.*, 2016). It was developed keeping antibody-antigen and enzyme-inhibitor interactions in mind. PatchDock algorithm carries out geometry based docking on the basis of shape complementarity. PatchDock algorithm has a relatively short run time. It can complete docking runs between 2 input proteins (of about 300 amino acids each) in less than 10 minutes on just a 1GHz processor. The web server serves as an interface for the PatchDock algorithm. During submission, the user may either upload the files on which the docking run has to be performed

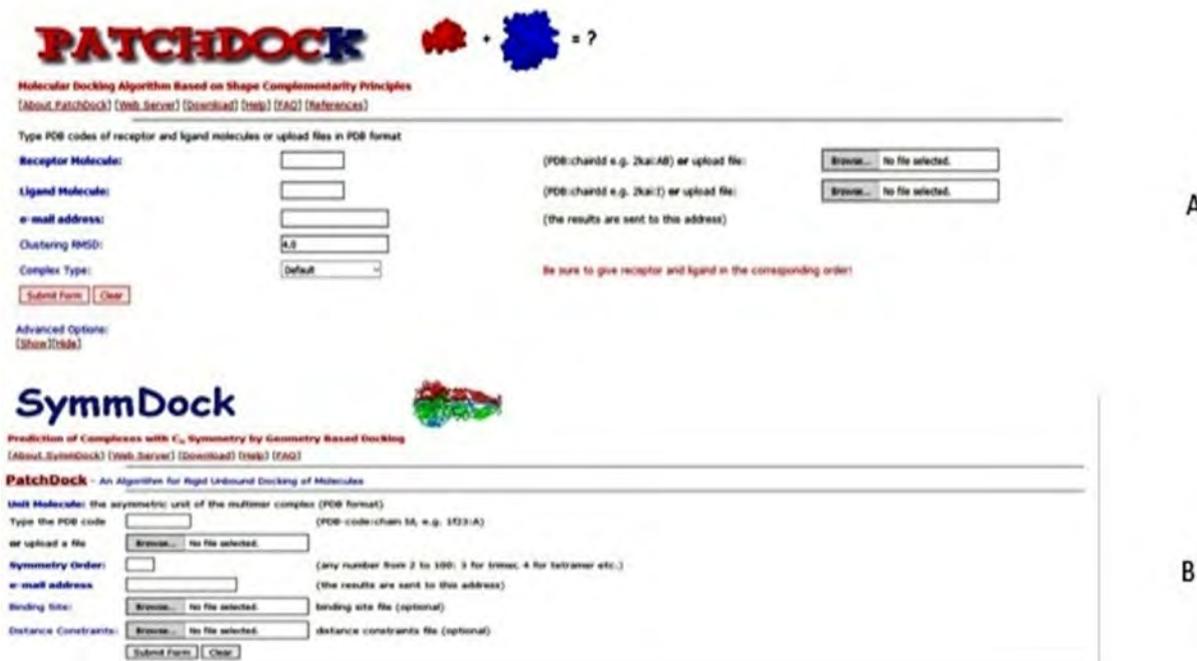


Figure 3: Snapshot showing (A) PatchDock web server interface and (B) SymmDock web server interface. Notice the similarity between the interfaces

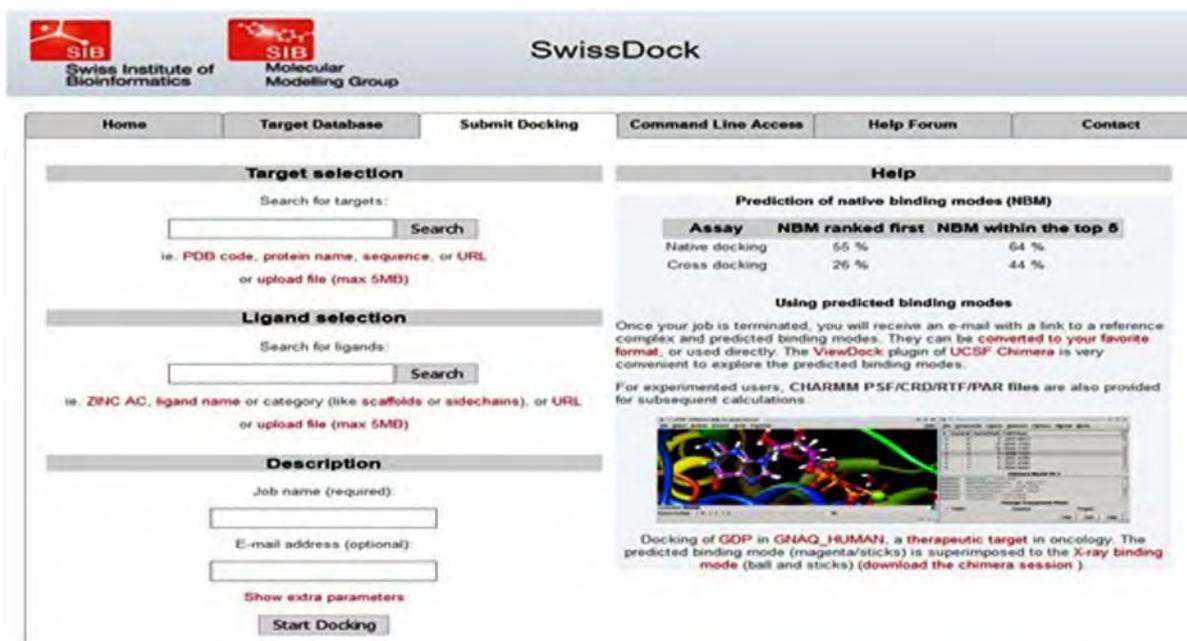


Figure 4: Snapshot showing SwissDock web server interface

in the PDB format or enter their PDB codes (Kumar *et al.*, 2015). The results as are sent to the user's email account upon completion of the docking run. The results with the top score can also be downloaded in a compressed file via a link on the results page. FiberDock web server is a useful tool for refining and ranking docking results from PatchDock.

### SymmDock

SymmDock web server uses the SymmDock algorithm for predicting the structure of homomulti-

mers which are cylindrically symmetrical. In addition to the PDB file of the molecule of interest, the user also has to enter the symmetry order of the molecule of interest. The user must keep in mind that SymmDock can only predict the quaternary structure of molecules with cyclic symmetry. The appearance of the SymmDock server is similar to PatchDock. The results are sent to the user via email (Schneidman *et al.*, 2005).

### FLIPDock

FLIPDock (Flexible Ligand Protein Docking) is molecular docking software developed by Yong Zhao

The screenshot shows the ClusPro web server interface. At the top, there are navigation tabs: Dock, Peptide Docking, Dimer Classification, Queue, Results, Papers, Help, and Contact. The main header features the ClusPro logo and the text 'protein-protein docking'. A 'sign\_out' link is visible in the top right corner. The central section is titled 'Dock' and contains a note: 'Note: all jobs by non logged in users will be publicly accessible. Please create an account if data is embargoed and needs to remain confidential'. Below this, there is a 'Job Name' input field and a 'Server' dropdown menu set to 'cpu'. A note specifies 'Accepted PDB Input: 20 standard amino acids and RNA (as receptor only), ref: RNA Select Heparin Mode to use Heparin as Ligand.' The interface is divided into 'Receptor' and 'Ligand' sections, each with 'PDB ID' and 'Chains' input fields and an 'Upload\_PDB' button. A note states: 'Whitespace separate desired chains. Leave chains blank to use all chains.' Below these fields is an 'Advanced Options' section with a checkbox and the text 'I agree to use ClusPro only for noncommercial purposes.' A 'Dock' button is at the bottom.

Figure 5: Snapshot showing ClusPro web server interface

The screenshot shows the YASARA Energy Minimization Server web interface. The title is 'YASARA Energy Minimization Server'. Below the title, there is a paragraph explaining the server's function: 'This server performs an energy minimization using the YASARA force field. Simply enter your email address, upload your protein model in PDB format and click the "Submit" button. Note that results will be placed in a public download area, do not submit confidential data. You can also use this functionality and much more on your own computer, it is part of YASARA Structure.' A citation is provided: 'Improving physical realism, stereochemistry, and side-chain accuracy in homology modeling: Four approaches that performed well in CASP8 Krieger E, Joo K, Lee J, Lee J, Raman S, Thompson J, Tyka M, Baker D, Karplus K Proteins. 2009;77 Suppl 9:114-22'. The main form area is titled 'YASARA Minimization Server' and contains input fields for 'Your email address', 'Either PDB ID', and 'or any other PDB file'. There are 'Browse...' and 'No file selected' buttons for the PDB file input, and a 'Submit...' button. Below the form, there are two bullet points: 'If you want to submit multiple structures, please wait until you received the confirmation email before submitting the next structure. Then you can sort the results based on the growing number in the download path and retrieve them in the proper order.' and 'If you don't get a confirmation email after submission, please check your spam folder and then wait 24 hours before reporting a problem, sometimes it takes longer.'

Figure 6: Snapshot showing YASARA minimization server web interface

and Michael Sanner at The Scripps Research Institute. It is coded in Python. It predicts docking poses between flexible ligands and flexible receptors. Although taking a flexible ligand into consideration during a docking run is computationally inexpensive, performing a docking run with a flexible receptor. FLIPDock utilizes a Flexibility Tree (FT) data structure in order to reduce the computational cost of using a flexible receptor (Zhao *et al.*, 2005).

### SwissDock

SwissDock server is a web server dedicated to protein-ligand docking simulations. It was developed by the Molecular Modeling group of The Swiss In-

stitute of Bioinformatics and is based on the docking program EADock DSS (Evolutionary algorithms Dock Dihedral Space Sampling). EADock DSS takes the best features from the highly accurate and flexible EADock 2 while being significantly faster than EADock. The protein and the ligand structure between which the docking needs to be carried out is submitted online. The SwissDock online interface is very user friendly and easy to use, allowing use by even beginners in protein-ligand docking studies. Moreover, since SwissDock is web server based, users do not have to worry about lack of computational resources for molecular docking as the SwissDock servers are utilized for docking. The results of the docking can be viewed from a URL provided upon submission. Alternatively, the user

Enter your email address [optional]:  Job name [optional]:

Copy and Paste one single structure in PDB format.

Perform post-refinement stereochemistry correction with MESH1 [Sample Input](#) [Clear Text Area](#)

[Click here to upload from your local computer](#)  
You may upload an archive (tar.gz, tar.bz2, zip) with multiple structures.

[Click here to upload one reference structure](#)  
CoRMSD, GDT-TS and GDT-HA will be calculated against this structure

### Using KoBa<sup>MIN</sup>

KoBa<sup>MIN</sup> makes available a very fast protein structure refinement protocol.

1. Paste one PDB file or upload a PDB file or an archive containing multiple structures.
2. Uncheck the MESH1 box for no stereochemistry correction.
3. Optionally, upload one reference structure for comparative GDT-HA, GDT-TS, and CoRMSD calculations.

**Figure 7: Snapshot showing KoBa<sup>MIN</sup> web server interface**

can state their email in order to receive the link to the results via email upon completion of the docking run. The predicted binding modes for the protein-ligand complex can be viewed online or downloaded as a zip file containing the PDB, DOCK and CHARMM format files. The files uploaded by the user and the results of the docking run are deleted within a period of 4 days (Grosdidier *et al.*, 2011).

### ClusPro

ClusPro web server, as previously mentioned, was the first molecular docking server made available to the scientific community. The user has to either upload the PDB file of the proteins of interest or enter their PDB code at the time of submission. The results are sent to the user's email upon completion of docking run. ClusPro carries out rigid body docking using the Fourier correlation method. ClusPro does not allow the receptor molecule to have more than 11999 atoms and does not permit the ligand to have more than 4700 atoms after energy minimization. In order to reduce the docking run time, users can use a perl script 'block.pl' (on the ClusPro web server) to restrict binding predictions to residues of interest on the receptor (Katchalski *et al.*, 1992). ClusPro also has symmetry functions which enable prediction of homomultimeric complex structures.

### Energy minimization tools

Energy minimization is the optimization of position of atoms in a molecule in order to attain a molecular structure with the lowest free energy. There is several of carrying out the structural refinement. Although comparative modeling does give correct backbone but it is inaccurate for side chains and H bonds (Bhattacharya *et al.*, 2013). Direct protein refinement can be carried out either by structural changes at global level or structural changes at the local level. Carrying out refinement

at the global level is more desirable but is significantly more computationally demanding. The latter does not give satisfactory results at the global level. Hence, a good refinement tool must achieve a balance between the two (Bhattacharya *et al.*, 2013).

### YASARA

YASARA (Yet Another Scientific Artificial Reality Application) is a molecular graphics, modelling and simulation software available on Linux, Windows, OS X and even Android. There are four stages in YASARA: view stage, model stage, dynamic stage and structure stage. Of these YASARA view is available for free as is. The remaining 3 require a license fee to be paid. Access to the other three can also be gained for free by contributing user side developments to the YASARA community. The YASARA minimization server is a web server for carrying out energy minimization of proteins structures. It is a part of YASARA structure and performs energy minimization with the help of YASARA force field (Krieger *et al.*, 2009).

Unlike the three stages of YASARA requiring a license fee, YASARA minimization server does not require any fee. It takes input in PDB format and emails the results to the user.

### KoBaMIN

KoBaMIN (Knowledge Based MINimization) web server is a freely available protein structure refinement and energy minimization web server with a simple and easy to use web interface. It does not require any registration and is totally free. KoBaMIN can also compare refined structure with a reference structure to determine the accuracy of the web server. The accuracy of any energy minimization depends on the accuracy of the force field on which the energy minimization tool is based on

Job name \*  Email

Copy and paste your initial structure [Example](#)

[Clear text](#)

Or upload structure from your local computer

No file selected.

Perform post refinement model analysis with:

MolProbity

RWPlus

### Job Submission

1. Provide job name and optional email id.
2. Copy-paste or upload initial structure.
3. Select optional post refinement analysis with MolProbity and/or RWPlus.
4. Submit the job for refinement.

### Retrieving Results

1. Web-based interactive status update.
2. Statistical and visual analysis of refinement.

**Figure 8: Snapshot showing 3D refine web server interface**

(Rodrigues *et al.*, 2013). Just like YASARA minimization server, KoBaMIN takes input in the PDB format. The results of the energy minimization are emailed to the user if the user submits their email during data submission. KoBaMIN also has the ability to take multiple structures if the structures to be refined are contained in a single zip, tar.gz or tar.bz2 archive. The KoBaMIN workflow involves 3 main steps: Validation of submitted structure, refinement or energy minimization of the submitted structure and obtaining the server output (Rodrigues *et al.*, 2013).

### 3D refine server

3D refine server is another openly accessible energy minimization server. The web interface of 3D refine is similar to KoBaMIN. 3D refine server workflow involves 3 main types: Validation of file type of submitted structure, H-bond optimization and energy minimization of optimized protein structure (Bhattacharya and Cheng, 2013). This workflow allows 3D refine to give improved results within a short period of time. 3D refine uses direct refinement of predicted model for structural prediction.

### CONCLUSION

Molecular docking and energy minimization software are an important part of the tool set of a researcher involved in computer aided drug design experiments. They are highly useful in predicting the interactions involving macromolecules as well

as their structures. Over the years, the accuracy, computational efficiency and accessibility of these programs have increased considerably. There are several free and open source tools for docking and structure refinement available online. Although their accuracy and computational efficiency has increased considerably, they are not without limitations. The main problem with docking tools is that they perform docking runs *in vacuo* (in vacuum) in most cases, leading to results which do not accurately depict the *in vivo* and *in vitro* conditions. As computational power increases, several docking tools have begun to appear which tackle this problem by, for instance, solvated docking. Another issue with docking tools, carrying out flexible docking, is being tackled by tools such as FLIPDock which simulate ligand and receptor flexibility. Energy minimization servers have allowed even beginners to carry out structural refinement by providing a user friendly interface and automatically setting appropriate values for advanced parameters. Such advances in docking and refinement have led and will continue to lead to greater productivity by researchers involved in structure based drug design and would also enable non experts to contribute to advancement of our knowledge in interactomics and drug design. We sincerely hope that this paper will be extremely useful for the researchers particularly from the developing countries where there is lack of funding for the research work and deadly diseases taking

the lives of so many people. In particular, drug development with the help of computationally techniques will be a bold step towards the drug designing process.

#### ACKNOWLEDGMENTS

Dr. Anish Kumar expresses a deep sense of gratitude to Mrs. Neera Rani and Mrs. Sheela Mahajan for constant encouragement provided by them through the research work.

**Conflict of Interests:** There are no conflicts of interest.

#### REFERENCES

Bhattacharya D, Cheng J. 3Drefine: Consistent Protein Structure Refinement by Optimizing Hydrogen Bonding Network and Atomic-Level Energy Minimization. *Proteins*, vol. 81, no 1, 2013, pp.119-131.

Bhattacharya D, Cheng J. i3Drefine software for protein 3D structure refinement and its assessment in CASP10. *PLoS One*.vol. 19, no. 8, 2013, e69648.

Carlson HA, McCammon JA. Accommodating protein flexibility in computational drug design. *Mol Pharmacol*, vol. 57, no.1, 2000, pp. 213-218.

Chaitanya M, Babajan B, Anuradha CM, Naveen M, Rajasekhar C, Madhusudana P, Kumar CS. Exploring the molecular basis for selective binding of Mycobacterium tuberculosis Asp kinase toward its natural substrates and feedback inhibitors: a docking and molecular dynamics study. *J Mol Model*,vol.16, no. 8, 2010, pp.1357-1367.

Clark KP, Ajay. Flexible ligand docking without parameter adjustment across four ligand-receptor complexes. *J Comput Chem*, vol.16, 1995, pp.1210-1226.

Comeau SR, Vajda S, Camacho CJ. Performance of the first protein docking server ClusPro in CAPRI rounds 3-5. *Proteins*, vol. 60, no. 2, 2005, pp. 239-244.

De Vries SJ, van Dijk M, Bonvin AM. The HADDOCK web server for data-driven biomolecular docking. *Nat Protoc*, vol.16, no. 8, 2010, pp.883-97.

Grosdidier A, Zoete V, Michielin O. Fast docking using the CHARMM force field with EADock DSS. *J Comput Chem*, vol. 32, 2011, pp. 2149-2159.

Hermann JC, Marti-Arbona R, Fedorov AA, Fedorov E, Almo SC, Shoichet BK, Raushel FM. Structure-based activity prediction for an enzyme of unknown function. *Nature*, vol. 448, 2007, pp.775-779.

Integrative Modeling of Biomolecular Complexes. *J Mol Biol*, vol.428. no.4, 2016, pp. 720-725.

Karaca E, Melquiond AS, de Vries SJ, Kastritis PL, Bonvin AM. Building macromolecular assemblies by information-driven docking: introducing the HADDOCK multibody docking server. *Mol Cell Proteomics*, vol. 9, no. 8, 2010, pp.1784-94.

Katchalski-Katzir E, Shariv I, Eisenstein M, Friesem AA, Aflalo C & Vakser IA. Molecular surface recognition: determination of geometric fit between proteins and their ligands by correlation techniques. *Proc Natl Acad Sci U S A*, vol. 89, no. 6, 1992, pp. 2195-2199.

Kolb P, Rosenbaum DM, Irwin JJ, Fung JJ, Kobilka B K, Shoichet BK. Structure-based discovery of  $\beta$ 2-adrenergic receptor ligands. *Proceedings of the National Academy of Sciences*, vol. 106, no 16, 2009, pp. 6843-6848.

Krieger E, Joo K, Lee J, et al. Improving physical realism, stereochemistry and side-chain accuracy in homology modeling: four approaches that performed well in CASP8. *Proteins*, vol.77, no.9, 2009, pp.114-122.

Kumar A, Ramanathan K. Exploring the structural and functional impact of the ALK F1174L mutation using bioinformatics approach. *J Mol Model*, vol.20, no.7, 2014, pp. 2324.

Kumar A, Ramanathan K. Virtual screening approach to identify potential ALK inhibitor from traditional Chinese medicine database. *RJPBCS*, vol.6, no.1, 2015, pp. 94-101.

Kumar A, Shanthi V, Ramanathan K. Computational investigation and experimental validation of crizotinib resistance conferred by C1156Y mutant anaplastic lymphoma kinase. *Mol Inform*, vol.34, no. 2-3, 2015, pp.105-114.

Kumar A, Shanthi V, Ramanathan K. Discovery of potential ALK inhibitors by virtual screening approach. *3 Biotech*, vol.6, 2016, pp.1-12.

Kumar A, Shanthi V, Ramanathan K. Management of crizotinib resistance in lung cancer using traditional plant source: An in silico strategies. *Biomedical Research*, vol.27, no.3, 2016, pp.794-800.

Kumar A, Shanthi V, Ramanathan K. QSAR and docking studies of Pubchem derivatives as potential ALK inhibitors for non-small cell lung cancer. *JOPCR*, vol. 8, no.9, 2016, pp.73-80.

Kumar A, Shanthi V, Ramanathan K. Structural and functional impact of G2032R mutation in ROS1 – a theoretical perspective. *AJPCR*, vol. 10, no.5, 2017, pp. 339-344.

- Laganà A, Veneziano D, Russo F. Computational design of artificial RNA molecules for gene regulation. *Methods Mol Biol*, vol.1269, 2015, pp.393-412.
- Lang PT, Brozell SR, Mukherjee S, Pettersen EF, Meng EC, Thomas V, Rizzo RC, Case DA, James TL, Kuntz ID. DOCK 6: Combining techniques to model RNA-small molecule complexes. *RNA*, vol.15, no.6, 2009, pp. 1219-1230.
- McInnes C. Virtual screening strategies in drug discovery. *Curr Opin Chem Biol*, vol. 11, 2007, pp.494-502.
- Ramachandran S, Kota P, Ding F, Dokholyan NV. Automated minimization of steric clashes in protein structures. *Proteins*, vol. 79, no.1, 2011, pp. 261-270.
- Rodrigues J, Levitt M, Chopra G. KoBaMIN: a knowledge-based minimization web server for protein structure refinement. *Nucleic Acids Res*, vol. 40, 2013, W323-W328.
- Schneidman D, Inbar Y, Nussinov R, Wolfson HJ. PatchDock and SymmDock: servers for rigid and symmetric docking. *Nucleic Acids Res*, vol. 33, 2005, pp.363-367.
- Seeliger D, de Groot BL. Ligand docking and binding site analysis with PyMOL and Auto-dock/Vina. *Journal of Computer-Aided Molecular Design*, vol.24, no.5, 2010, pp. 417-422.
- Teague SJ. Implications of protein flexibility for drug discovery. *Nat Rev Drug Discov*, vol. 2, 2003, pp. 527-541.
- Trott O, Olson AJ. AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization and multi-threading. *J Comput Chem*, vol.31, no. 2, 2010, pp. 455-461.
- Tuszynska I, Magnus M, Jonak K, Dawson W, Bujnicki JM. NPDock: a web server for protein-nucleic acid docking. *Nucleic Acids Res*, vol.43, 2015, pp. 425-430.
- Van Zundert GC, Rodrigues JP, Trellet M, Schmitz C, Kastiris PL, Karaca E, Melquiond AS, van Dijk M, de Vries SJ, Bonvin AM. The HADDOCK2.2 Web Server: User-Friendly
- Zhao Y, Sanner M. FLIPDock: Docking flexible ligands into flexible receptors. *Proteins*, vol.68, 2007, pp.726-737.