Antifungal potential of Fenugreek Seeds (Trigonella foenum-graecum) Crude Extracts against *Microsporum gypseum*

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**ABSTRACT**

*Trigonella foenum-graecum* (*fabaceae*) is commonly used as condiments and spices in Indian and Asian food to flavour, colour, and texture of food, and it is employed in various medicinal purposes in traditional systems. The biological activity of fenugreek can be easily accessed from previous research conducted by several researchers. The present research was conducted to find out the antifungal potential of various extracts of dried powder of fenugreek seeds by means of paper disc diffusion method, with petroleum ether, ethyl acetate, ethanol, and aqueous solvents in 25, 50, and 100 concentrations against *Microsporum gypseum*. Clotrimazole was used as a standard. The present study revealed that fenugreek is a potent antifungal agent against *Microsporum gypseum*. The ethanol extract of fenugreek using 100 concentrations depicted the highest zone of inhibition of 16.510+ 0.85mm and 38.395% of mycelial inhibition against a tested pathogen. While drug extracts in other solvents also revealed reasonable to least antifungal potential. This finding tells us that fenugreek extracts tested proved to be a potent antifungal agent against *Microsporum gypseum*. It was found that ethanol extract of fenugreek is best effective against tested strain. This exploration of fenugreek extracts has confirmed its importance, particularly in the area of influence on dermatophytic fungal strain.

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**INTRODUCTION**

The microbial and fungal infections are becoming crucial trouble to mankind, and they are the foremost reason of morbidity and mortality of many developing countries (Ahmed *et al.*, 2012). Many antimicrobial agents are presently accessible for the treatment and management of infectious diseases (Karuppiah and Rajaram, 2012). In order to overcome the ill effects and resistance caused due to synthetic drugs, the World Health Organization have motivated many researchers to exploit natural products for their great therapeutic potential (Talebi *et al.*, 2014). A huge variety of herbal antifungal agents derived from traditional medicinal plants are existing for the treatment of dermatophytes (Kimm *et al.*, 2015). In the present scenario, medicinal plants and their phytoconstituents are gaining attention owing to the fact that herbal drugs are lesser in cost, easily accessible, and with fewer or no side effects (Malik *et al.*, 2015). *Trigonella foenum graecum* (Fenugreek) belong the family...
Fabaceae is one of the ancient traditional medicinal plants and has a long history of its therapeutic uses (King et al., 2015). It contains lysine and L-tryptophan rich proteins, mucilaginous fibre and other rare chemical constituents such as saponins, coumarin, fenugreek, nicotinic acid, sapogenin-sphytic acid, scopoletin and trigonelle which has therapeutic effects. (Billaud and , 2001).

**MATERIALS AND METHODS**

**Preparation of plant extract**

Fenugreek dried seeds were purchased from the local market for the preparation of the extract. The dried seeds of methi were grounded to form a powder with the help of a mechanical grinder. Herbarium sheet is submitted with the Pharmacognosy department Chandigarh college of Pharmacy—Landran Mohali with voucher no. CCP/TFG/069. Extracts were prepared by extracting methi seed powder successively with petroleum ether, ethyl acetate, ethanol, and aqueous solvents and tested against Microsporum gypseum. The prepared extract was weighed and stored in airtight sample bottles. The filtered extracts were tested against dermatophytes at three different concentrations viz. 25 μml, 50 μml, and 100 μml.

**Procedure and Procurement of strain**

The antifungal potential of an extract of dried seeds of methi was evaluated by the Paper disc diffusion method. The test organisms used were the dermatophyte strains of Microsporum gypseum, which was procured from IMTECH, Chandigarh. MTCC No. was 2829. Sabouraud Dextrose agar was used as a culture media according to the manufacturer’s direction. The dermatophyte cultures were aseptically inoculated on Sabouraud agar plate and subjected to incubation at 28 ºC for 72 hours. The antifungal potential was determined by measuring the zone of inhibition (ZOI) around the discs and percentage inhibition after the period of incubation (Rawal and Adhikari, 2016).

**Data Analysis**

Data from antifungal screening was analyzed with the help of simple statistics from Microsoft Excel and recorded in appropriate tables as a mean ± standard deviation of the mean.

**RESULTS AND DISCUSSION**

Antifungal potential of extracts of seeds of Trigonella foenum-graecum against the tested fungal strain Microsporum gypseum can be seen in Table 1. The Pet ether extract of methi showed 5.89 mm ZOI at 25 μml concentration. 50 μml concentrations were moderately effective with 10.62 mm zone of inhibition. At 100 μml, the zone of inhibition was observed to be as 12.85 mm. The ethyl acetate extract showed a 10.44 mm inhibition zone at 25 μml concentration. 50 μml concentrations were effective with 13.69 mm inhibition zone. 15.89 mm inhibition zone was observed at 100 μml. The ethanol extract showed a 12.42 mm inhibition zone at 25 μml concentration. 50 μml concentrations were moderately effective with 14.32 mm inhibition zone. At 100 μml, the inhibition zone was observed to be as 16.51 mm. While its aqueous extract showed a 3.88 mm inhibition zone at 25 μml concentration. 50 μml concentrations were effective with a 9.74 mm inhibition zone. 10.12 mm inhibition zone was observed at 100 μml concentration. The antifungal potential was determined by comparing the activity of extracts with the Clotrimazole, in which the zone of inhibition was 43mm. Percentage inhibition was also calculated, which was 38.395 % with 100 μml ethanol extract depicted in Table 2. Many important phytoconstituents obtained from plants, having antifungal and antidermatophytic properties, is of vital significance to medicinal treatments (Bondad-Reantaso et al., 2005). The results of this study showed that of all the extracts screened, Trigonella foenum-graecum ethanol extract had higher inhibitory activ-
Table 1: Mean Zone of Inhibition in different solvents (mm) of the *Trigonella foenum-graecum*

<table>
<thead>
<tr>
<th>Crude Drug</th>
<th>Concentration (µl)</th>
<th>Mean Zone of Inhibition in different solvents (mm)</th>
<th>Pet Ether</th>
<th>Ethyl acetate</th>
<th>Ethanol</th>
<th>Aqueous</th>
<th>Clotrimazole</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Trigonella foenum-graecum</em></td>
<td>25 µl</td>
<td>5.891+ 0.58</td>
<td>10.443+ 0.67</td>
<td>12.425+ 0.65</td>
<td>3.889+ 1.20</td>
<td>43mm</td>
<td></td>
</tr>
<tr>
<td></td>
<td>50 µl</td>
<td>10.622+ 0.65</td>
<td>13.690+ 0.86</td>
<td>14.320+ 0.75</td>
<td>9.740+ 0.72</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td></td>
<td>100 µl</td>
<td>12.852+ 0.55</td>
<td>15.890+ 0.75</td>
<td>16.510+ 0.85</td>
<td>10.120+ 0.41</td>
<td>-</td>
<td></td>
</tr>
</tbody>
</table>

Table 2: Percentage inhibition (%) of various extracts of the *Trigonella foenum-graecum*

<table>
<thead>
<tr>
<th>Crude Drug</th>
<th>Concentration (µl)</th>
<th>Percentage inhibition (%)</th>
<th>Pet Ether</th>
<th>Ethyl acetate</th>
<th>Ethanol</th>
<th>Aqueous</th>
<th>Clotrimazole</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>50 µl</td>
<td>23.995 31.197</td>
<td>32.676 21.924</td>
<td>-</td>
<td>21.924 -</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>100 µl</td>
<td>29.883 36.953</td>
<td>38.395 23.534</td>
<td>-</td>
<td>23.534 -</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 3: Phytochemical tests of the extracts of the *Trigonella foenum-graecum*

<table>
<thead>
<tr>
<th>Chemical compound</th>
<th>Petroleum extract</th>
<th>Ethyl acetate</th>
<th>Ethanol extract</th>
<th>Aqueous extract</th>
<th>N-Absent, P-Present</th>
</tr>
</thead>
<tbody>
<tr>
<td>Coumarins</td>
<td>N</td>
<td>N</td>
<td>P</td>
<td>P</td>
<td>N-Absent</td>
</tr>
<tr>
<td>Alkaloids</td>
<td>P</td>
<td>P</td>
<td>N</td>
<td>N</td>
<td>P-Present</td>
</tr>
<tr>
<td>Tannins</td>
<td>N</td>
<td>N</td>
<td>P</td>
<td>P</td>
<td>N-Absent</td>
</tr>
<tr>
<td>Terpenoids</td>
<td>N</td>
<td>N</td>
<td>P</td>
<td>P</td>
<td>N-Absent</td>
</tr>
<tr>
<td>Saponins</td>
<td>N</td>
<td>N</td>
<td>P</td>
<td>P</td>
<td>N-Absent</td>
</tr>
<tr>
<td>Flavonoids</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>P</td>
<td>N-Absent</td>
</tr>
</tbody>
</table>

CONCLUSIONS

The present research provides evidence about the antifungal potential of crude extracts of *Trigonella foenum-graecum* seeds against *Microsporum gypseum*. The antifungal potential is different depending on the polarity of the solvent utilized in the extraction process. From the study, it can be depicted that ethanol extract of methi seeds are promising as compared to other solvents. Furthermore, quantitative phytochemical analysis can be conducted in the future to isolate and identify the phytoconstituents liable for the antifungal potential.

REFERENCES


